

Review

Oseltamivir against Influenza in Severe Acute Respiratory Infection (SARI): Review

¹Tingre G, ²Dhakane R

¹Department of Zoology, Nowrosjee Wadia College, Pune, Maharashtra, India.

²Department of Microbiology, Jayawantrao Sawant College of Commerce and Science, Pune, Maharashtra, India.

Article Info

Article history:

Received: August 20, 2020

Accepted: October 26, 2020

Published: November 1, 2020

Keywords: Severe Acute Respiratory Infection, oseltamivir, prophylaxis, influenza.

Corresponding Author:

Gayatri Tingre,

Email: gayatritingre22@gmail.com

Abstract

Many viral diseases have been generating potential health issues to humans. Severe Acute Respiratory Infection (SARI), a disease of respiratory system, is one of them. Treatment of this disease is crucial factor to save human life using oseltamivir because it has been used by medical practitioners and received promising results. Diverse medicines are being investigated for the same purpose. In this review, we have examined the oseltamivir which is used against the infection in question for its efficiency.

©Author(s). This work is licensed under a [Creative Commons Attribution-NonCommercial-ShareAlike 4.0 International License](https://creativecommons.org/licenses/by-nc-sa/4.0/) that permits noncommercial use of the work provided that credit must be given to the creator and adaptation must be shared under the

1. Introduction:

In humanoid olden times, there were several cases of virus-related pandemic situations including polio [1], Ebola [2], smallpox [3], chickenpox [4] and HIV-AIDS [5] that placed hominoid life on severe hazard [3]. On the other hand, such sicknesses were eradicated via serum [6], cures [7] and prediction [8]. The stated infections were life-threatening and caused prospective biotic damage in the globe [9,10,11]. Of them, aerial and infectious illnesses [12] are extra difficult for people, and their prevention is challenging [13]. Viral communicable infections cause major illnesses and death in humans [14].

Lower respiratory contagions are the lethal transmissible infections, producing 3.2 million deaths universally in 2015 [14]. To fight with virus-related

illnesses, serums and antiviral medications have been used mutually [14]. The acute respiratory contagions are the principal reason of deceases in early life stages all over the world [15] and give rise to 1.9 million deceases each year [16], of which 70 % occur in developing nations [17]. Additionally, it was responsible for 30% of all childhood deaths in the developing world [18].

Severe acute respiratory infection (SARI) is one of the principal reasons of deaths between young stages of life globally [19]. SARIs are termed as an acute respiratory illness of current arrival (in seven days) expressed by fever ($\geq 38^{\circ}\text{C}$), cough and breathlessness necessitating hospitalization [20]. SARI is caused by both influenza and other viruses [21,22, 23], for example, respiratory syncytial virus [21,23],

Parainfluenza subtypes 1, 2 along with 3 [21,22], bocavirus [22], Parainfluenza virus type 3 [22, 94], Piconavirus (EV/RV) [22], Adenovirus [19,21,22,23], Influenza A (H1N1) [22], Rhinovirus [19], Influenza B [93], Human metapneumovirus (hMPV) [22] and bacteria [21].

Hence, the effective treatment strategy of SARI becomes essential. In this review, we have investigated the oseltamivir drug for its efficiency against the influenza infection, one of the characters of SARI, since it has been used by medical practitioners and received promising results.

2. Treatment:

Treating influenza infection in SARI is one of the challenges in the medical fields. The judgement given by the clinician is the basis of criteria for the admission in hospital as well as treatment in general, and the admissions in hospitals are not always connected with severity [24]. Hence, employing a case definition is important [24]. In comparison, no scientific evidence of treatment against SARI was reported, and also, scientific case definition of SARI has not been provided.

The usual diagnosis of SARIs is conducted clinically and antibiotics are used for its treatment as per bacterial culture along with susceptibility tests with care that supports in the case where virus diagnosis facilities are unavailable [19]. Although, no scientific proof of specific antiviral drugs against SARI has been provided and according to Xuan C et al. [25] and Loeffelholz M et al. [26], antiviral drug oseltamivir is used against influenza but its properties and clinical applications against SARI has not been provided. Generally, the antiviral drug oseltamivir is used against various viral infections. GS 4104 (oseltamivir, TAMIFLU) has appeared as a guaranteed antiviral treatment as well as prophylaxis of infection by influenza in humans [77].

3. Properties and clinical applications of oseltamivir:

Oseltamivir (Tamiflu) is an antiviral drug which acts against influenza A and B [27]. To add, it creates obstruction in the function of viral neuraminidase protein [27]. Neuraminidase bearing influenza virus with reduced oseltamivir carboxylate sensitivity changed characters in vitro, and is compromised as far

as infection is concerned in addition to the capacity of replication in vivo [43]. There are some inhibitors that can be used to control viral infection by creating barrier in the life processes of virus.

In this sequence, neuraminidase inhibitors are able to reduce the symptoms caused by influenza including fever, head ache and myalgias within approximately 0.7 to 1.5 days if their administration occurred before 48 hours after infection [27] by inhibiting the viral release from cell (figure 1). In accordance with the current guidelines to treat pregnant women suffering from the influenza infection, one of the permitted neuraminidase inhibitors can be used [33].

Interestingly, viral neuraminidase of influenza A and B are selectively inhibited by oseltamivir carboxylate [27]. The production of oseltamivir was important success in medical field [27]. A fat soluble side chain of the active drug attaches to the neuraminidase enzyme, interrupting its capability to cleave sialic acid residues on the outer layer of the infected cell, causing an incapability to release progeny virus particles [28,29,30], hence viral natural life cycle gets disturbed.

After the infection by influenza virus, oseltamivir administrations are not able to interrupt the cellular immune responses [31]. Unfortunately, the Tamiflu has good clinical tolerance except insignificant gastrointestinal upset [32] and therefore, more research work is required in the field of drug designing to engineer Tamiflu in such a way that gastrointestinal upset can be tolerated.

4. Pharmacokinetics and distribution:

As administration of oseltamivir in human body is considered, its fate in the body should be studied. Additionally, in the blood, gastrointestinal tract along with the liver, oseltamivir ethyl ester that is to be administered orally is absorbed in a good way in addition to its cleavage with esterases speedily [27,33]. Then, it is finally converted to its active form oseltamivir carboxylate which is equally distributed all over the body, together with the upper and lower respiratory tract [27].

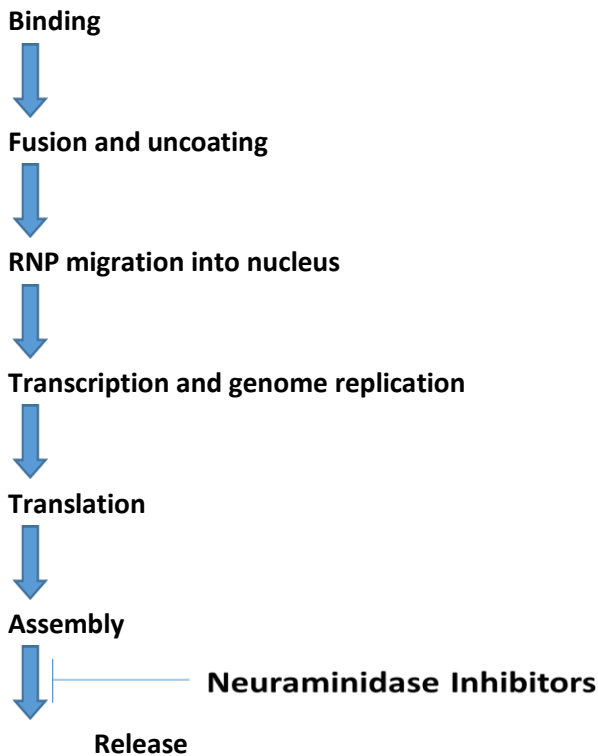


Figure 1: Target sites of the antiviral agents. Note: Reprinted from Beigel J I and Bray M [70] and modified by Tingre, G., & Dhakane, R. (2020).

5. Route of administration:

Oseltamivir can be delivered orally only in the form 75 mg, 45 mg and 30 mg tablets and white tutti-frutti flavoured suspension [33]. 75 mg two times a day for 5 days is approved dose of the medicine under study for adults [33].

6. Dosage:

The prescribed dosing of a medicine is very important to cure the disease successfully. Nevertheless, since active metabolite is observed in increased concentration, there is no requirement of the dosage adjustment [35]. Oe et al. [38] reported that it is compulsory to adjust the dose with help of increasing 2 mg/kg two times daily because the clearance rate of metabolite which was active was faster in younger than adults. This indicates that there are opposite opinions among the researchers regarding dosage adjustment of oseltamivir and hence, this area demands further scientific investigation.

Likewise, the drug is contraindicated among younger owing to its toxicity [38] which may be serious issue among the patients with respect of their health to treat disease. The combined efforts are

recommended to be taken from global researchers to resolve the appeared contraindication and make the drug safe for use by younger patients. The common dose of oseltamivir can be lowered to 75 mg one time a day in order to achieve treatment as well as either 75 mg each other day or 30 mg suspension everyday for prophylaxis when a patient has a creatinine clearance of <30 mL/min [33]. Besides, doses are recommended to be given after hemodialysis [33].

However, in case of pregnant women, some studies indicated the requirement of higher doses of oseltamivir (75 mg TID) while other studies suggest that there was no need of adjustment of dose [63, 64, 65]. Making the treatment dose of oseltamivir double, the patients who are hospitalized does not show virologic efficacy increase with exception for infections by influenza B or clinical effectiveness even if one ICU based RCT suggested that making the standard dose triple was connected with the increase of clearance of viral RNA from the tract of respiratory system [66, 67, 68].

7. Prophylaxis:

The drug oseltamivir showed promising results when used practically in the patients suffering from the influenza infection. If administered in the time less than 48 hours, it showed affectivity against the infection under study [27]. 33 volunteers were treated with the drug in question (8/21 volunteers) or placebo (8/12 volunteers) for 5 days with a dose as 100 mg orally [27]. As well, it is considered to prevent the influenza infection in the patients of more than or equal to one year with dosing pattern as once in a day [33]. What is more, the effectiveness of oseltamivir with dose as 75 mg once daily for 6 weeks in the process of preventing influenza infection in healthy as well as non-immunized adult was reported as 84%, and it was 50% in the process of preventing infections by influenza regardless of symptoms [34]. In the residents of immunized nursing home, the prophylaxis efficacy was 92% against ill health as compared to placebo [39].

In a prophylaxis study of household-contact, fairly lower efficacy was observed [40]. Further, approximately 80% protective efficacy was reported in relation with seasonal prophylaxis of patients with

high-risk immune compromising pattern against influenza illness which is confirmed by RT-PCR [41]. Fortunately, Hayden et al. 1999 [42] examined not only safety but also efficiency of oseltamivir in the adults with the ages in the range of 18-40 years. Authors reported that 74% of all volunteers were protected by oseltamivir against influenza A [42].

As a conclusion, both trials indicated that in adults, oseltamivir was not only safe but also effective to prevent the influenza infection. The risk of developing influenza was reduced by 70-90 % showed by meta-analysis study of seven prevention trials [37]. However, since the prophylaxis results into the resistant mutants, the precaution should be taken while prescribing oseltamivir in patients who exposed with an index case for prophylaxis [44]. As a result, generally, monitoring is required in these cases [33].

8. Chemoprophylaxis:

Unfortunately, there is unavailability of reference to administer oseltamivir before infection prophylaxis by H1N1 for the influenza virus of the swine [27]. As soon as confirmation of H1N1 infection, chemoprophylaxis should be provided and continued for 10 days subsequent to the last diagnosis [27].

9. Merits:

In order to alleviate the non-complicated influenza illness, oseltamivir (75mg two times a day for 5 days) makes the time short with reduction of illness severity, fever duration, time required to reach at normal activity, viral shedding quantity, impaired activity duration, complications resulted due to antibiotic use especially bronchitis, compared to placebo in earlier healthy adults [45, 46, 47]. Also, oseltamivir showed effectiveness up to 72 hours after onset of symptom in children in Bangladesh [48]. The paediatric studies that included children of 2 weeks old showed that oseltamivir was not only safe but also it reduced duration and illness significantly along with the time required to resume the complete activities as well as existence of complications due to antibiotic use [49-53].

Furthermore, observational studies provided the major part of existing literature based on the efficacy as well as safety of oseltamivir in high-risk or elderly persons involving those who were suffered from

immunodeficiency or cardiopulmonary conditions [54-57]. Among such hospitalized and high risk members, there was a benefit if antiviral therapy started at least 5 days and within 48 hours after onset of symptoms [58]. Likewise, the studies of adults who are hospitalized suggest that the early therapy reduced the occurrence of complications of lower respiratory tract, duration of illness, ICU-level care requirement and duration of mortality and shedding [45, 46, 56, 57, 59]. This clearly admits the value of early treatment of the infection to eradicate it easily.

10. Adverse reactions:

Oseltamivir is not exceptional among the drugs that show side effects on the body. It is connected with abdominal discomfort, nausea and emesis (less often) in the minority of the treated patients [33]. To add, in oseltamivir recipients, both vomiting and nausea exist at about 10-15 % higher frequency [33]. In contrary to this, the reports of post marketing show that oseltamivir may be connected not often with the skin rash, thrombocytopenia or dysfunction [33]. However, rash, swelling of face or tongue, epidermal necrolysis which is toxic, tests of abnormal liver function, hepatitis, seizures, arrhythmias, confusion aggravation of diabetes are other reported adverse effects [69]. Thus, the adverse effects of oseltamivir cause health concern to the patients.

Besides, there are reports of not only abnormal neurologic but also behavioural characters that have hardly ever resulted into deaths among mostly children and many of these reports arrived from Japan [33]. The pregnant women infected by influenza got clear therapeutic benefit due to oseltamivir which is safe [60, 61, 62]. The available data suggest that such events are secondary to infections by influenza than oseltamivir therapy [71, 72]. Besides, steady-state pharmacokinetics of generally used immunosuppressive agents is not affected by oseltamivir [73].

Additionally, cimetidine inhibitor of cytochrome P450 has no effect on the plasma levels of oseltamivir carboxylate or oseltamivir [27]. With no any modification in dose, renal excretion rate of oseltamivir may be controlled with help of Probenecid [74]. Oseltamivir might impair immunogenicity of

concurrent intranasal influenza vaccine which is live attenuated [33]. Unless the potential advantage justifies the risk (potential) to the foetus, the oseltamivir prescription in the pediatric patients with ages less than 1 year should be avoided [75].

11. Resistance:

Developing resistance against medicaments is a natural process of life. Likewise, many strains of H1N1 influenza A virus responsible for pandemic were sensitive to neuraminidase inhibitors i.e oseltamivir as well as zanamivir [27]. Oseltamivir resistance is connected with NA mutations E119, H274Y, R292K and R152K [76]. Even if the R292K neuraminidase mutation give resistance at high level to GS 4071 (neuraminidase inhibitor) in vitro, its effect on the virulence by viruses render this mutation with limited clinical significance [78]. Following oseltamivir phosphate treatment, the H274Y mutation found in the influenza A/H1N1 neuraminidase active site leaves the viruses compromised severely both not only in vivo but also in vitro [79].

The NA inhibitors show their activity against many influenza A and B viruses and cause lesser side effects, however, the generation of resistance against oseltamivir by various influenza viruses has been reported [81]. For example, with rare incidences of limited transmission, H1N1 virus infections which are sporadic have been reported in 2009 [80,83,84,85]. In reality, H1N1 virus strains which are oseltamivir resistant emerged within 48 hours after starting of treatment [86].

The rare occurrence of spread of influenza B virus strains that are resistant to oseltamivir or 2009 H1N1 virus strains received from persons treated by oseltamivir was reported and clinical isolates with lowered susceptibility to zanamivir were obtained irregularly from immunocompromised children after

prolonged therapy [87, 88, 89, 90]. During the treatment of influenza which is seasonal, the development of resistance against oseltamivir or zanamivir has been detected [91,92]. For management of the disease, the data related with variables connected with the disease was obtained including requirement of non-invasive and invasive mechanical ventilation, vasoactive drugs, corticosteroid use (inhaled or systemic), prescription of antibiotic compounds and antiviral oseltamivir [93].

12. Conclusion:

Oseltamivir is a suitable drug to treat influenza infection that occurs in the patients who suffer from Severe Acute Respiratory Infection (SARI) with exception of some adverse effects. Future research may include investigation of oseltamivir effect against viruses other than influenza which are involved in SARI although individual treatments against them were practiced with no aim to treat SARI.

Authors' contributions:

GT: Wrote manuscript. RD: Developed an idea and verified the data.

Competing interest:

Authors declare that no competing interest exists among them.

Ethical Statement:

Since this is a review article, no ethical permission is required.

Grant Support Details:

The authors have not received funding for this work from any agency.

References:

1. Chumakov, K., Ehrenfeld, E., Wimmer, E., & Agol, V. I. (2007). Vaccination against polio should not be stopped. *Nature reviews. Microbiology*, 5(12), 952–958. <https://doi.org/10.1038/nrmicro1769>
2. Malvy, D., McElroy, A. K., de Clerck, H., Günther, S., & van Griensven, J. (2019). Ebola virus disease. *Lancet (London, England)*, 393(10174),936–948. [https://doi.org/10.1016/S0140-6736\(18\)33132-5](https://doi.org/10.1016/S0140-6736(18)33132-5)
3. Malvy, D., McElroy, A. K., de Clerck, H., Günther, S., & van Griensven, J. (2019). Ebola virus disease. *Lancet (London, England)*, 393(10174),936–948. [https://doi.org/10.1016/S0140-6736\(18\)33132-5](https://doi.org/10.1016/S0140-6736(18)33132-5)
4. Heininger, U., & Seward, J. F. (2006). Varicella. *Lancet (London, England)*, 368(9544), 1365–1376. [https://doi.org/10.1016/S0140-6736\(06\)69561-5](https://doi.org/10.1016/S0140-6736(06)69561-5)
5. A, K., S, C., RB, S., H, V., SS, S., & A, G. (2016). A REVIEW ON: HIV AIDS. *Indian Journal of Pharmaceutical and Biological Research*, 4(03), 69-73. <https://doi.org/10.30750/ijpbr.4.3.9>

6. Bandyopadhyay, A. S., Garon, J., Seib, K., & Orenstein, W. A. (2015). Polio vaccination: past, present and future. *Future microbiology*, 10(5),791–808. <https://doi.org/10.2217/fmb.15.19>
7. Ogra, P. L., & Volovitz, B. (1987). Diagnosis and treatment of viral infections. *Bulletin of the New York Academy of Medicine*, 63(6), 475–483.
8. Coffin, J. M. (1999). *Molecular biology of HIV*. In: *The Evolution of HIV* (Crandall K.A., ed.). pp. 3-40. Johns Hopkins University Press, Baltimore, MD.
9. Hu, B., Zeng, L. P., Yang, X. L., Ge, X. Y., Zhang, W., Li, B., Xie, J. Z., Shen, X. R., Zhang, Y. Z., Wang, N., Luo, D. S., Zheng, X. S., Wang, M. N., Daszak, P., Wang, L. F., Cui, J., & Shi, Z. L. (2017). Discovery of a rich gene pool of bat SARS-related coronaviruses provides new insights into the origin of SARS coronavirus. *PLoS pathogens*, 13(11), e1006698. <https://doi.org/10.1371/journal.ppat.1006698>
10. Dudas, G., Carvalho, L. M., Rambaut, A., & Bedford, T. (2018). MERS-CoV spillover at the camel-human interface. *eLife*, 7,e31257. <https://doi.org/10.7554/eLife.31257>
11. Leopardi, S., Holmes, E. C., Gastaldelli, M., Tassoni, L., Priori, P., Scaravelli, D., Zamperin, G., & De Benedictis, P. (2018). Interplay between co-divergence and cross-species transmission in the evolutionary history of bat coronaviruses. *Infection, genetics and evolution : journal of molecular epidemiology and evolutionary genetics in infectious diseases*, 58, 279–289. <https://doi.org/10.1016/j.meegid.2018.01.012>
12. Wang, N., Li, S. Y., Yang, X. L., Huang, H. M., Zhang, Y. J., Guo, H., Luo, C. M., Miller, M., Zhu, G., Chmura, A. A., Hagan, E., Zhou, J. H., Zhang, Y. Z., Wang, L. F., Daszak, P., & Shi, Z. L. (2018). Serological Evidence of Bat SARS-Related Coronavirus Infection in Humans, China. *Virologica Sinica*, 33(1),104–107. <https://doi.org/10.1007/s12250-018-0012-7>
13. Dhakane, R.D., Shinde, A.H, Bhattacharjee, S.B., & Wagh, S.G. (2020). Can Coronaviridae Viruses Reappear with their Novel Variants in Upcoming Years? A Review. *International Journal of Microbial Science*, 1(1), 1-5.
14. Takizawa, N., & Yamasaki, M. (2017). Current landscape and future prospects of antiviral drugs derived from microbial products. *The Journal of antibiotics*, 71(1), 45–52. Advance online publication. <https://doi.org/10.1038/ja.2017.115>
15. Li, H., Wei, Q., Tan, A., & Wang, L. (2013). Epidemiological analysis of respiratory viral etiology for influenza-like illness during 2010 in Zhuhai, China. *Virology journal*, 10, 143. <https://doi.org/10.1186/1743-422X-10-143>
16. Parveen, S., Sullender, W. M., Fowler, K., Lefkowitz, E. J., Kapoor, S. K., & Broor, S. (2006). Genetic variability in the G protein gene of group A and B respiratory syncytial viruses from India. *Journal of clinical microbiology*, 44(9), 3055–3064. <https://doi.org/10.1128/JCM.00187-06>
17. Williams, B. G., Gouws, E., Boschi-Pinto, C., Bryce, J., & Dye, C. (2002). Estimates of world-wide distribution of child deaths from acute respiratory infections. *The Lancet. Infectious diseases*, 2(1), 25–32. [https://doi.org/10.1016/s1473-3099\(01\)00170-0](https://doi.org/10.1016/s1473-3099(01)00170-0)
18. Bharaj, P., Sullender, W. M., Kabra, S. K., Mani, K., Cherian, J., Tyagi, V., Chahar, H. S., Kaushik, S., Dar, L., & Broor, S. (2009). Respiratory viral infections detected by multiplex PCR among pediatric patients with lower respiratory tract infections seen at an urban hospital in Delhi from 2005 to 2007. *Virology journal*, 6, 89. <https://doi.org/10.1186/1743-422X-6-89>
19. Malhotra, B., Swamy, M. A., Janardhan Reddy, P. V., & Gupta, M. L. (2016). Viruses causing severe acute respiratory infections (SARI) in children ≤5 years of age at a tertiary care hospital in Rajasthan, India. *The Indian journal of medical research*, 144(6),877–885. https://doi.org/10.4103/ijmr.IJMR_22_15
20. World Health Organization (2013). Regional Office for Europe. Overview of sentinel systems for hospitalized severe acute respiratory infections (SARI) represented in the weekly Euro Flu surveillance bulletin (as of 10 February 2013). Available from: http://www.euro.who.int/_data/assets/pdf_file/0005/186863/Overview-of-SARI-Surveillance-Systemsfinal.pdf accessed on May 5, 2013.
21. Pan American Health Organization (2009). Regional Office of the World Health Organization. Health Establishments Preparation for Unusual or Unexpected Cases or Clusters of Severe Acute Respiratory Infection (SARI). Available from: https://www.paho.org/hq/dmdocuments/2009/SARI%20English_Module_Final.pdf
22. Zhang, C., Zhu, N., Xie, Z., Lu, R., He, B., Liu, C., Ma, X., & Tan, W. (2013). Viral etiology and clinical profiles of children with severe acute respiratory infections in China. *PLoS one*, 8(8), e72606. <https://doi.org/10.1371/journal.pone.0072606>
23. Hatem, A., Mohamed, S., Abu Elhassan, U. E., Ismael, E., Rizk, M. S., El-Kholy, A., & El-Harras, M. (2019). Clinical characteristics and outcomes of patients with severe acute respiratory infections (SARI): results from the Egyptian surveillance study 2010-2014. *Multidisciplinary respiratory medicine*, 14,11. <https://doi.org/10.1186/s40248-019-0174-7>
24. Meerhoff, T. J., Simaku, A., Ulqinaku, D., Torosyan, L., Gribkova, N., Shimanovich, V., Chakhunashvili, G., Karseladze, I., Yesmagambetova, A., Kuatbayeva, A., Nurmatov, Z., Otorbaeva, D., Lupulescu, E., Popovici, O., Smorodintseva, E., Sominina, A., Holubka, O., Onyshchenko, O., Brown, C. S., & Gross, D. (2015). Surveillance for severe acute respiratory infections (SARI) in hospitals in the WHO European region - an exploratory analysis of risk factors for a severe outcome in influenza-positive SARI cases. *BMC infectious diseases*, 15, 1. <https://doi.org/10.1186/s12879-014-0722-x>
25. Xuan, C., Yan, L., & Zegang, W. (2013). Rapid detection of acute respiratory virus and atypical bacteria infections in

- children. *Jundishapur J Microbiol*, Online ahead of Print, 6(5):6236. doi: [10.5812/ijm.6236](https://doi.org/10.5812/ijm.6236)
26. Loeffelholz, M., & Chonmaitree, T. (2010). Advances in diagnosis of respiratory virus infections. *International journal of microbiology*, 2010, 126049. <https://doi.org/10.1155/2010/126049>
 27. Sharma Prince, Roy Ram, Chaudhary, Anurag. (2010). Neuraminidase inhibitors: Oseltamivir, Peramivir, Synthesis and Profile. *Journal of Pharmacy Research*, 3.
 28. McNicholl, I. R., & McNicholl, J. J. (2001). Neuraminidase inhibitors: zanamivir and oseltamivir. *The Annals of pharmacotherapy*, 35(1),57–70. <https://doi.org/10.1345/aph.10118>
 29. Bardsley-Elliott, A., & Noble, S. (1999). Oseltamivir. *Drugs*, 58(5),851–862. <https://doi.org/10.2165/00003495-199958050-00007>
 30. Lew, W., Chen, X., & Kim, C. U. (2000). Discovery and development of GS 4104 (oseltamivir): an orally active influenza neuraminidase inhibitor. *Current medicinal chemistry*, 7(6),663–672. <https://doi.org/10.2174/0929867003374886>
 31. Burger, R. A., Billingsley, J. L., Huffman, J. H., Bailey, K. W., Kim, C. U., & Sidwell, R. W. (2000). Immunological effects of the orally administered neuraminidase inhibitor oseltamivir in influenza virus-infected and uninfected mice. *Immunopharmacology*, 47(1),45–52. [https://doi.org/10.1016/s0162-3109\(99\)00184-8](https://doi.org/10.1016/s0162-3109(99)00184-8)
 32. Doucette, K. E., & Aoki, F. Y. (2001). Oseltamivir: a clinical and pharmacological perspective. *Expert opinion on pharmacotherapy*, 2(10),1671–1683. <https://doi.org/10.1517/14656566.2.10.1671>
 33. Ison, M. G., & Hayden, F. G. (2017). Antiviral Agents Against Respiratory Viruses. *Infectious Diseases*, 1318–1326.e2. <https://doi.org/10.1016/B978-0-7020-6285-8.00154-4>
 34. McClellan, K., & Perry, C. M. (2001). Oseltamivir: a review of its use in influenza. *Drugs*, 61(2), 263–283. <https://doi.org/10.2165/00003495-200161020-00011>
 35. He, G., Massarella, J., & Ward, P. (1999). Clinical pharmacokinetics of the prodrug oseltamivir and its active metabolite Ro 64-0802. *Clinical pharmacokinetics*, 37(6), 471–484. <https://doi.org/10.2165/00003088-199937060-00003>
 36. Rayner, C. R., Bulik, C. C., Kamal, M. A., Reynolds, D. K., Toovey, S., Hammel, J. P., Smith, P. F., Bhavnani, S. M., Van Wart, S. A., Ambrose, P. G., & Forrest, A. (2013). Pharmacokinetic-pharmacodynamic determinants of oseltamivir efficacy using data from phase 2 inoculation studies. *Antimicrobial agents and chemotherapy*, 57(8), 3478–3487. <https://doi.org/10.1128/AAC.02440-12>
 37. Cooper, N. J., Sutton, A. J., Abrams, K. R., Wailoo, A., Turner, D., & Nicholson, K. G. (2003). Effectiveness of neuraminidase inhibitors in treatment and prevention of influenza A and B: systematic review and meta-analyses of randomised controlled trials. *BMJ (Clinical research ed.)*, 326(7401), 1235. <https://doi.org/10.1136/bmj.326.7401.1235>
 38. Oo, C., Barrett, J., Hill, G., Mann, J., Dorr, A., Dutkowski, R., & Ward, P. (2001). Pharmacokinetics and dosage recommendations for an oseltamivir oral suspension for the treatment of influenza in children. *Paediatric drugs*, 3(3), 229–236. <https://doi.org/10.2165/00128072-200103030-00005>
 39. Kamali, A., & Holodniy, M. (2013). Influenza treatment and prophylaxis with neuraminidase inhibitors: a review. *Infection and drug resistance*, 6, 187–198. <https://doi.org/10.2147/IDR.S36601>
 40. Welliver, R., Monto, A. S., Carewicz, O., Schatteman, E., Hassman, M., Hedrick, J., Jackson, H. C., Huson, L., Ward, P., Oxford, J. S., & Oseltamivir Post Exposure Prophylaxis Investigator Group (2001). Effectiveness of oseltamivir in preventing influenza in household contacts: a randomized controlled trial. *JAMA*, 285(6), 748–754. <https://doi.org/10.1001/jama.285.6.748>
 41. Ison, M. G., Szakaly, P., Shapira, M. Y., Kriván, G., Nist, A., & Dutkowski, R. (2012). Efficacy and safety of oral oseltamivir for influenza prophylaxis in transplant recipients. *Antiviral therapy*, 17(6), 955–964. <https://doi.org/10.3851/IMP2192>
 42. Hayden, F. G., Treanor, J. J., Fritz, R. S., Lobo, M., Betts, R. F., Miller, M., Kinnersley, N., Mills, R. G., Ward, P., & Straus, S. E. (1999). Use of the oral neuraminidase inhibitor oseltamivir in experimental human influenza: randomized controlled trials for prevention and treatment. *JAMA*, 282(13),1240–1246. <https://doi.org/10.1001/jama.282.13.1240>
 43. Carr, J., Ives, J., Kelly, L., Lambkin, R., Oxford, J., Mendel, D., Tai, L., & Roberts, N. (2002). Influenza virus carrying neuraminidase with reduced sensitivity to oseltamivir carboxylate has altered properties in vitro and is compromised for infectivity and replicative ability in vivo. *Antiviral research*, 54(2), 79–88. [https://doi.org/10.1016/s0166-3542\(01\)00215-7](https://doi.org/10.1016/s0166-3542(01)00215-7)
 44. Baz, M., Abed, Y., Papenburg, J., Bouhy, X., Hamelin, M. E., & Boivin, G. (2009). Emergence of oseltamivir-resistant pandemic H1N1 virus during prophylaxis. *The New England journal of medicine*, 361(23), 2296–2297. <https://doi.org/10.1056/NEJMc0910060>
 45. Fiore, A. E., Fry, A., Shay, D., Gubareva, L., Bresee, J. S., Uyeki, T. M., & Centers for Disease Control and Prevention (CDC) (2011). Antiviral agents for the treatment and chemoprophylaxis of influenza --- recommendations of the Advisory Committee on Immunization Practices (ACIP). *MMWR. Recommendations and reports: Morbidity and mortality weekly report. Recommendations and reports*, 60(1), 1–24.
 46. Hsu, J., Santesso, N., Mustafa, R., Brozek, J., Chen, Y. L., Hopkins, J. P., Cheung, A., Hovhannisyan, G., Ivanova, L., Flottorp, S. A., Saeterdal, I., Wong, A. D., Tian, J., Uyeki, T. M., Akl, E. A., Alonso-Coello, P., Smaill, F., & Schünemann, H. J. (2012). Antivirals for treatment of influenza: a systematic review and meta-analysis of observational studies. *Annals of*

- internal medicine*, 156(7), 512–524. <https://doi.org/10.7326/0003-4819-156-7-20120403000411>
47. Kaiser, L., Wat, C., Mills, T., Mahoney, P., Ward, P., & Hayden, F. (2003). Impact of oseltamivir treatment on influenza-related lower respiratory tract complications and hospitalizations. *Archives of internal medicine*, 163(14), 1667–1672. <https://doi.org/10.1001/archinte.163.14.1667>
 48. Fry, A. M., Goswami, D., Nahar, K., Sharmin, A. T., Rahman, M., Gubareva, L., Azim, T., Bresee, J., Luby, S. P., & Brooks, W. A. (2014). Efficacy of oseltamivir treatment started within 5 days of symptom onset to reduce influenza illness duration and virus shedding in an urban setting in Bangladesh: a randomised placebo-controlled trial. *The Lancet. Infectious diseases*, 14(2), 109–118. [https://doi.org/10.1016/S1473-3099\(13\)70267-6](https://doi.org/10.1016/S1473-3099(13)70267-6)
 49. Acosta, E. P., Jester, P., Gal, P., Wimmer, J., Wade, J., Whitley, R. J., Kimberlin, D. W., & National Institute of Allergy and Infectious Diseases Collaborative Antiviral Study Group (2010). Oseltamivir dosing for influenza infection in premature neonates. *The Journal of infectious diseases*, 202(4), 563–566. <https://doi.org/10.1086/654930>
 50. Kamal, M. A., Acosta, E. P., Kimberlin, D. W., Gibiansky, L., Jester, P., Niranjana, V., Rath, B., Clinch, B., Sánchez, P. J., Ampofo, K., Whitley, R., & Rayner, C. R. (2014). The posology of oseltamivir in infants with influenza infection using a population pharmacokinetic approach. *Clinical pharmacology and therapeutics*, 96(3), 380–389. <https://doi.org/10.1038/clpt.2014.120>
 51. Kimberlin, D. W., Acosta, E. P., Prichard, M. N., Sánchez, P. J., Ampofo, K., Lang, D., Ashouri, N., Vanchiere, J. A., Abzug, M. J., Abughali, N., Caserta, M. T., Englund, J. A., Sood, S. K., Spigarelli, M. G., Bradley, J. S., Lew, J., Michaels, M. G., Wan, W., Cloud, G., Jester, P., ... National Institute of Allergy and Infectious Diseases Collaborative Antiviral Study Group (2013). Oseltamivir pharmacokinetics, dosing, and resistance among children aged <2 years with influenza. *The Journal of infectious diseases*, 207(5), 709–720. <https://doi.org/10.1093/infdis/jis765>
 52. Whitley, R. J., Hayden, F. G., Reisinger, K. S., Young, N., Dutkowski, R., Ipe, D., Mills, R. G., & Ward, P. (2001). Oral oseltamivir treatment of influenza in children. *The Pediatric infectious disease journal*, 20(2), 127–133. <https://doi.org/10.1097/00006454-200102000-00002>
 53. Piedra, P. A., Schulman, K. L., & Blumentals, W. A. (2009). Effects of oseltamivir on influenza-related complications in children with chronic medical conditions. *Pediatrics*, 124(1), 170–178. <https://doi.org/10.1542/peds.2008-0977>
 54. Kumar, D., Michaels, M. G., Morris, M. I., Green, M., Avery, R. K., Liu, C., Danziger-Isakov, L., Stosor, V., Estabrook, M., Gantt, S., Marr, K. A., Martin, S., Silveira, F. P., Razonable, R. R., Allen, U. D., Levi, M. E., Lyon, G. M., Bell, L. E., Huprikar, S., Patel, G., ... American Society of Transplantation H1N1 Collaborative Study Group (2010). Outcomes from pandemic influenza A H1N1 infection in recipients of solid-organ transplants: a multicentre cohort study. *The Lancet. Infectious diseases*, 10(8), 521–526. [https://doi.org/10.1016/S1473-3099\(10\)70133-X](https://doi.org/10.1016/S1473-3099(10)70133-X)
 55. Reid, G., Huprikar, S., Patel, G., Razonable, R. R., Mossad, S., Levi, M., Gregg, K., Shoham, S., Humar, A., Adams, W., & Kumar, D. (2013). A multicenter evaluation of pandemic influenza A/H1N1 in hematopoietic stem cell transplant recipients. *Transplant infectious disease: an official journal of the Transplantation Society*, 15(5), 487–492. <https://doi.org/10.1111/tid.12116>
 56. Lee, N., & Ison, M. G. (2012). Diagnosis, management and outcomes of adults hospitalized with influenza. *Antiviral therapy*, 17(1 Pt B), 143–157. <https://doi.org/10.3851/IMP2059>
 57. Lee, N., & Ison, M. G. (2012). Editorial commentary. "Late" treatment with neuraminidase inhibitors for severely ill patients with influenza: better late than never?. *Clinical infectious diseases : an official publication of the Infectious Diseases Society of America*, 55(9), 1205–1208. <https://doi.org/10.1093/cid/cis642>
 58. Louie, J. K., Yang, S., Acosta, M., Yen, C., Samuel, M. C., Schechter, R., Guevara, H., & Uyeki, T. M. (2012). Treatment with neuraminidase inhibitors for critically ill patients with influenza A (H1N1)pdm09. *Clinical infectious diseases : an official publication of the Infectious Diseases Society of America*, 55(9), 1198–1204. <https://doi.org/10.1093/cid/cis636>
 59. Muthuri, S. G., Venkatesan, S., Myles, P. R., Leonardi-Bee, J., Al Khuwaitir, T. S., Al Mamun, A., Anovadiya, A. P., Azziz-Baumgartner, E., Báez, C., Bassetti, M., Beovic, B., Bertisch, B., Bonmarin, I., Booy, R., Borja-Aburto, V. H., Burgmann, H., Cao, B., Carratala, J., Denholm, J. T., Dominguez, S. R., ... Nguyen-Van-Tam, J. S. (2014). Effectiveness of neuraminidase inhibitors in reducing mortality in patients admitted to hospital with influenza A H1N1pdm09 virus infection: a meta-analysis of individual participant data. *The Lancet. Respiratory medicine*, 2(5), 395–404. [https://doi.org/10.1016/S2213-2600\(14\)70041-4](https://doi.org/10.1016/S2213-2600(14)70041-4)
 60. Beau, A. B., Hurault-Delarue, C., Vial, T., Montastruc, J. L., Damase-Michel, C., & Lacroix, I. (2014). Safety of oseltamivir during pregnancy: a comparative study using the EFEMERIS database. *BJOG : an international journal of obstetrics and gynaecology*, 121(7), 895–900. <https://doi.org/10.1111/1471-0528.12617>
 61. Wollenhaupt, M., Chandrasekaran, A., & Tomianovic, D. (2014). The safety of oseltamivir in pregnancy: an updated review of post-marketing data. *Pharmacoepidemiology and drug safety*, 23(10), 1035–1042. <https://doi.org/10.1002/pds.3673>
 62. Saito, S., Minakami, H., Nakai, A., Unno, N., Kubo, T., & Yoshimura, Y. (2013). Outcomes of infants exposed to oseltamivir or zanamivir in utero during pandemic (H1N1) 2009. *American journal of obstetrics and gynecology*, 209(2), 130.e1–130.e1309. <https://doi.org/10.1016/j.ajog.2013.04.007>

63. Beigi, R. H., Han, K., Venkataramanan, R., Hankins, G. D., Clark, S., Hebert, M. F., Easterling, T., Zajicek, A., Ren, Z., Mattison, D. R., Caritis, S. N., & Obstetric-Fetal Pharmacology Research Units Network (2011). Pharmacokinetics of oseltamivir among pregnant and nonpregnant women. *American journal of obstetrics and gynecology*, 204(6 Suppl 1), S84–S88. <https://doi.org/10.1016/j.ajog.2011.03.002>
64. Greer, L. G., Leff, R. D., Rogers, V. L., Roberts, S. W., McCracken, G. H., Jr, Wendel, G. D., Jr, & Sheffield, J. S. (2011). Pharmacokinetics of oseltamivir according to trimester of pregnancy. *American journal of obstetrics and gynecology*, 204(6 Suppl 1), S89–S93. <https://doi.org/10.1016/j.ajog.2011.03.005>
65. Greer, L. G., Leff, R. D., Rogers, V. L., Roberts, S. W., McCracken, G. H., Jr, Wendel, G. D., Jr, & Sheffield, J. S. (2011). Pharmacokinetics of oseltamivir according to trimester of pregnancy. *American journal of obstetrics and gynecology*, 204(6 Suppl 1), S89–S93. <https://doi.org/10.1016/j.ajog.2011.03.005>
66. Lee, N., Hui, D. S., Zuo, Z., Ngai, K. L., Lui, G. C., Wo, S. K., Tam, W. W., Chan, M. C., Wong, B. C., Wong, R. Y., Choi, K. W., Sin, W. W., Lee, E. L., Tomlinson, B., Hayden, F. G., & Chan, P. K. (2013). A prospective intervention study on higher-dose oseltamivir treatment in adults hospitalized with influenza A and B infections. *Clinical infectious diseases : an official publication of the Infectious Diseases Society of America*, 57(11),1511–1519. <https://doi.org/10.1093/cid/cit597>
67. South East Asia Infectious Disease Clinical Research Network (2013). Effect of double dose oseltamivir on clinical and virological outcomes in children and adults admitted to hospital with severe influenza: double blind randomised controlled trial. *BMJ (Clinical research ed.)*, 346, f3039. <https://doi.org/10.1136/bmj.f3039>
68. Kumar A. (2013). Viral Clearance with Standard or Triple Dose Oseltamivir Therapy in Critically Ill Patients with Pandemic (H1N1) 2009 Influenza. ICAAC 2013. *Denver, Colorado*, 2013:B-1470.
69. FDA–Food & Drug Administration, FDA Approves Tamiflu for Prevention of Influenza in Children Under Age 12. Available at: <http://test.fda.gov/NewsEvents/Newsroom/PressAnnouncements/2005/ucm1045252.htm>
70. Beigel, J., & Bray, M. (2008). Current and future antiviral therapy of severe seasonal and avian influenza. *Antiviral research*, 78(1),91–102. <https://doi.org/10.1016/j.antiviral.2008.01.003>
71. Toovey, S., Prinssen, E. P., Rayner, C. R., Thakrar, B. T., Dutkowski, R., Koerner, A., Chu, T., Sirzen-Zelenskaya, A., Britschgi, M., Bansod, S., & Donner, B. (2012). Post-marketing assessment of neuropsychiatric adverse events in influenza patients treated with oseltamivir: an updated review. *Advances in therapy*, 29(10), 826–848. <https://doi.org/10.1007/s12325-012-0050-8>
72. Hoffman KB, Demakas A, Erdman CB, Dimbil M, Doraiswamy PM. Neuropsychiatric adverse effects of oseltamivir in the FDA Adverse Event Reporting System, 1999-2012. *BMJ*. 2013 Jul 23;347:f4656. [doi: 10.1136/bmj.f4656](https://doi.org/10.1136/bmj.f4656) PMID: 23881998
73. Lam, H., Jeffery, J., Sitar, D. S., & Aoki, F. Y. (2011). Oseltamivir, an influenza neuraminidase inhibitor drug, does not affect the steady-state pharmacokinetic characteristics of cyclosporine, mycophenolate, or tacrolimus in adult renal transplant patients. *Therapeutic drug monitoring*, 33(6), 699–704. <https://doi.org/10.1097/FTD.0b013e3182399448>
74. Hill, G., Cihlar, T., Oo, C., Ho, E. S., Prior, K., Wiltshire, H., Barrett, J., Liu, B., & Ward, P. (2002). The anti-influenza drug oseltamivir exhibits low potential to induce pharmacokinetic drug interactions via renal secretion-correlation of in vivo and in vitro studies. *Drug metabolism and disposition: the biological fate of chemicals*, 30(1), 13–19. <https://doi.org/10.1124/dmd.30.1.13>
75. Schirmer, P., & Holodniy, M. (2009). Oseltamivir for treatment and prophylaxis of influenza infection. *Expert opinion on drug safety*, 8(3), 357–371. <https://doi.org/10.1517/14740330902840519>
76. McKimm-Breschkin, J., Trivedi, T., Hampson, A., Hay, A., Klimov, A., Tashiro, M., Hayden, F., & Zambon, M. (2003). Neuraminidase sequence analysis and susceptibilities of influenza virus clinical isolates to zanamivir and oseltamivir. *Antimicrobial agents and chemotherapy*, 47(7), 2264–2272. <https://doi.org/10.1128/aac.47.7.2264-2272.2003>
77. Lew, W., Chen, X., & Kim, C. U. (2000). Discovery and development of GS 4104 (oseltamivir): an orally active influenza neuraminidase inhibitor. *Current medicinal chemistry*, 7(6),663–672. <https://doi.org/10.2174/0929867003374886>
78. Tai, C. Y., Escarpe, P. A., Sidwell, R. W., Williams, M. A., Lew, W., Wu, H., Kim, C. U., & Mendel, D. B. (1998). Characterization of human influenza virus variants selected in vitro in the presence of the neuraminidase inhibitor GS 4071. *Antimicrobial agents and chemotherapy*, 42(12), 3234–3241. <https://doi.org/10.1128/AAC.42.12.3234>
79. Ives, J. A., Carr, J. A., Mendel, D. B., Tai, C. Y., Lambkin, R., Kelly, L., Oxford, J. S., Hayden, F. G., & Roberts, N. A. (2002). The H274Y mutation in the influenza A/H1N1 neuraminidase active site following oseltamivir phosphate treatment leave virus severely compromised both in vitro and in vivo. *Antiviral research*, 55(2), 307–317. [https://doi.org/10.1016/s0166-3542\(02\)00053-0](https://doi.org/10.1016/s0166-3542(02)00053-0)
80. Yen, H. L., Herlocher, L. M., Hoffmann, E., Matrosovich, M. N., Monto, A. S., Webster, R. G., & Govorkova, E. A. (2005). Neuraminidase inhibitor-resistant influenza viruses may differ substantially in fitness and transmissibility. *Antimicrobial agents and chemotherapy*, 49(10),4075–4084. <https://doi.org/10.1128/AAC.49.10.4075-4084.2005>
81. Centres for Disease Control and Prevention (CDC) [Internet]. Antiviral drug resistance among influenza viruses. Seasonal

- influenza (flu) health professionals antiviral drugs. c2015 [cited 2016 September 25]. Available from: <http://www.cdc.gov/flu/professionals/antivirals/antiviral-drug-resistance.htm>
82. Centres for Disease Control and Prevention (CDC) [Internet]. Antiviral drug resistance among influenza viruses. Seasonal influenza (flu) health professionals antiviral drugs. c2015 [cited 2016 September 25]. Available from: <http://www.cdc.gov/flu/professionals/antivirals/antiviral-drug-resistance.htm>
 83. Le, Q. M., Wertheim, H. F., Tran, N. D., van Doorn, H. R., Nguyen, T. H., Horby, P., & Vietnam H1N1 Investigation Team (2010). A community cluster of oseltamivir-resistant cases of 2009 H1N1 influenza. *The New England journal of medicine*, 362(1),86–87. <https://doi.org/10.1056/NEJMc0910448>
 84. Centers for Disease Control and Prevention (CDC) (2009). Oseltamivir resistant novel influenza A (H1N1) virus infection in two immunosuppressed patients-Seattle, Washington, 2009. *MMWR*, 58(32):893.
 85. Centers for Disease Control and Prevention (CDC) (2009). Oseltamivir resistant 2009 pandemic influenza A (H1N1) virus infection in two summer campers receiving prophylaxis-north Carolina-2009. *MMWR*, 58(35):969–972.
 86. Inoue M, Barkham T, Leo YS, Chan KP, Chow A, Wong CW, Tze Chuen Lee R, Maurer-Stroh S, Lin R, Lin C. Emergence of oseltamivir-resistant pandemic (H1N1) 2009 virus within 48 hours. *Emerg Infect Dis*. 2010 Oct; 16(10):1633-6. [doi: 10.3201/eid1610.100688](https://doi.org/10.3201/eid1610.100688) PMID: 20875299; PMCID: PMC3294403.
 87. Le, Q. M., Wertheim, H. F., Tran, N. D., van Doorn, H. R., Nguyen, T. H., Horby, P., & Vietnam H1N1 Investigation Team (2010). A community cluster of oseltamivir-resistant cases of 2009 H1N1 influenza. *The New England journal of medicine*, 362(1),86–87. <https://doi.org/10.1056/NEJMc0910448>
 88. Hatakeyama, S., Sugaya, N., Ito, M., Yamazaki, M., Ichikawa, M., Kimura, K., Kiso, M., Shimizu, H., Kawakami, C., Koike, K., Mitamura, K., & Kawaoka, Y. (2007). Emergence of influenza B viruses with reduced sensitivity to neuraminidase inhibitors. *JAMA*, 297(13),1435–1442. <https://doi.org/10.1001/jama.297.13.1435>
 89. Gubareva, L. V., Matrosovich, M. N., Brenner, M. K., Bethell, R. C., & Webster, R. G. (1998). Evidence for zanamivir resistance in an immunocompromised child infected with influenza B virus. *The Journal of infectious diseases*, 178(5), 1257–1262. <https://doi.org/10.1086/314440>
 90. Ison, M. G., Gubareva, L. V., Atmar, R. L., Treanor, J., & Hayden, F. G. (2006). Recovery of drug-resistant influenza virus from immunocompromised patients: a case series. *The Journal of infectious diseases*, 193(6), 760–764. <https://doi.org/10.1086/500465>
 91. Centers for Disease Control and Prevention (CDC) (2009). Oseltamivir resistant 2009 pandemic influenza A (H1N1) virus infection in two summer campers receiving prophylaxis-north Carolina-2009. *MMWR*, 58(35):969–972.
 92. Stephenson, I., Democratis, J., Lackenby, A., McNally, T., Smith, J., Pareek, M., Ellis, J., Birmingham, A., Nicholson, K., & Zambon, M. (2009). Neuraminidase inhibitor resistance after oseltamivir treatment of acute influenza A and B in children. *Clinical infectious diseases: an official publication of the Infectious Diseases Society of America*, 48(4), 389–396. <https://doi.org/10.1086/596311>
 93. Fica, A., Sotomayor, V., Fasce, R., Dabanch, J., Soto, A., Charpentier, P., Guerrero, G., Olivares, F., Triantafilo, V., Omeiri, N. E., & Gainza-Lein, M. (2019). Severe acute respiratory infections (SARI) from influenza in adult patients in Chile: the experience of a sentinel hospital. *Revista panamericana de salud publica = Pan American journal of public health*, 43, e1. <https://doi.org/10.26633/RPSP.2019.1>
 94. Pretorius, M. A., Tempia, S., Walaza, S., Cohen, A. L., Moyes, J., Variava, E., Dawood, H., Seleka, M., Hellferscee, O., Treurnicht, F., Cohen, C., & Venter, M. (2016). The role of influenza, RSV and other common respiratory viruses in severe acute respiratory infections and influenza-like illness in a population with a high HIV sero-prevalence, South Africa 2012-2015. *Journal of clinical virology : the official publication of the Pan American Society for Clinical Virology*, 75,21–26. <https://doi.org/10.1016/j.jcv.2015.12.004>

Cite this article as: Tingre G, Dhakane R. Oseltamivir against Influenza in Severe Acute Respiratory Infection (SARI): Review. *International Journal of Microbial Science* [Internet]. 2020;1(1). Available from: <http://dx.doi.org/10.55347/theijms.v1i1.2>